

# A neuroprotective potential of berberine in Acrylamide induced neurotoxicity

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#### **ABSTRACT: -**

Neurotoxicity refers to the harmful effects on the nervous system, including its normal functions like neuronal transmission, connectivity, and survival. Various substances, such as drugs, toxins, and food additives, can induce neurotoxicity. Acrylamide (ACR), also known as 2-propenamide, is a colorless, odorless, water-soluble crystalline solid with a low molecular weight. It has been widely synthesized for many years. One of the primary culprits of neurotoxicity is oxidative stress and mitochondrial dysfunction. Acrylamide toxicity impacts both the peripheral and central nervous systems. Berberine, an isoquinoline alkaloid derived from different Berberis species, is frequently used in the treatment of neurological and neurodegenerative disorders like Alzheimer's, Huntington's disease (HD), Parkinson's disease, mental depression, schizophrenia, anxiety, and certain stroke conditions. Berberine is a small molecule that easily crosses the blood-brain barrier (BBB). Numerous clinical and preclinical studies have demonstrated the positive effects of berberine on a range of health issues, including metabolic, neurological, and cardiovascular problems. This review seeks to outline the potential neuroprotective properties of berberine against ACR-induced neurotoxicity.

Keywords: - neurotoxicity, berberine, acrylamide, mitochondrial dysfunction, oxidative stress.

#### I. **INTRODUCTION: -**

Neurotoxicity is the result of harmful agents affecting the normal functioning of the nervous system, including processes like neuronal transmission, connectivity, and survival. Various substances, including drugs, toxins, and food additives, can potentially induce neurotoxicity [1]. Both experimental research in animals and observations in industrial settings involving humans have indicated that Acrylamide (ACR)

toxicity has detrimental effects on both the peripheral and central nervous systems. Human exposure to ACR can occur through inhalation, ingestion (via the diet), and skin absorption, leading to neurological symptoms [2]. The neurotoxic impact of ACR is progressive, affecting both the peripheral and central nervous systems, with cumulative damage observed. Repeated exposure to ACR can initially manifest with mild symptoms that can progress to severe impairment and irreversible harm [3].

ACR, also known as 2-propenamide, is a crystalline solid characterized by its colorless, odorless, and water-soluble properties. Figure No. 01 Shows the Physico-Chemical Properties of ACR. This low-molecular-weight substituted alkene has been produced extensively for many years and is commonly used as an intermediate in organic compound manufacturing and as a monomer in polyacrylamide production [4-5]. ACR also finds applications in the cosmetics and textile industries, laboratories, and as a soil conditioner in wastewater treatment [6-8]. A significant source of ACR is the cooking of carbohydrate-rich foods with asparagine and low sugar content at temperatures above 120°C and with limited moisture [9-10]. Dietary intake is the primary source of exposure for the general population, though exposure from cosmetics can also contribute. The estimated chronic daily dietary intake for the average person ranges from 0.3 to 0.7 grams per kilogram of body weight (g/kg BTW) [11-12]. Common dietary sources of ACR include French fries (averaging 308 g/kg), potato chips (averaging 389 g/kg), bread (averaging 42 g/kg), cookies (averaging 265 g/kg), and coffee (averaging 522 g/kg in dry coffee). Notably, ACR has also been found in significant amounts in cigarette smoke, ranging from 497 to 169 nanograms per cigarette [13]. Children's ACR intake tends to increase with age, peaking between



12 and 18 years, reflecting greater consumption of fast-food items like potato chips or French fries [11-12].Various regulatory bodies have set permissible limits for ACR in drinking water, with the World Health Organization (WHO) at 1 gram per liter (g/l), the US Environmental Protection Agency at 0.5 g/l, and the European Union at 1 g/l. Occupational exposure limits for ACR in ambient air have been established at 0.3 milligrams per cubic meter (mg/m3) for time-weighted averages of 8 hours (Occupational Safety and Health Administration) or 10 hours (National Institute for Occupational Safety and Health) [14-15]. The discovery of ACR in food prompted numerous international conferences, bringing together scientists to assess the implications of these findings. Regulatory agencies such as the US Federal FDA, the UK Foods Standard Agency, Health Canada, and the Swedish National Food Administration all issued statements following these discussions. The broad spectrum of foods potentially containing ACR residues and the prospect of eliminating specific items from diets

raise concerns regarding health risks associated with dietary choices. In addition, foodborne illnesses impact millions annually, causing thousands of fatalities. Over 200 research projects have been initiated to gain a better understanding of the risks associated with ACR exposure in humans (EFSA, 2005; FAO/WHO, 2005). In early 2005, a WHO/FAO meeting further examined these findings in the context of food safety (JECFA, 2005). The Risk Assessment section of this document includes some of the meeting's remarks and recommendations [16].Berberine is a natural alkaloid compound found in various plants, including goldenseal, barberry, and Oregon grape. It has been traditionally used in various traditional medicine practices, particularly in Chinese and Ayurvedic medicine, for its potential health benefits. Berberine has gained attention in modern research due to its diverse pharmacological properties, including Neuroprotective Effects, Antiinflammatory, Antimicrobial, and Antioxidant effects.

S.NO.	CLASS	PROPERTIES			
1.	Molecular formula	C <sub>3</sub> H <sub>5</sub> NO			
2.	IUPAC Name	2-propenamide			
3.	Molecular Weight	71.078			
	[g/mol]				
4.	Physical characteristics	White, Crystalline solid at room			
		temperature			
5.	Density	30 °C 1.127 g/cm3			
6.	Melting Point	84 - 84.5 °C			
7.	Boiling Point	125 °C at 3.3 Pa			
8.	Vapour Pressure	0.9 Pa at 25 °C			
9.	log POW	0.67 to 1.65			
10.	Water Solubility	2.155 g/l at 30 °C			

Table	01: - Pl	hysico-	Chemical	Properties

# Factors responsible for ACR toxicity: -

One of the primary factors contributing to ACR toxicity is its formation during certain cooking processes. ACR is produced when carbohydrate-rich foods, such as potatoes and cereals, are cooked at high temperatures, typically above 120°C, in the presence of amino acids, particularly asparagine, and reducing sugars. Common cooking methods like frying, baking, and roasting can trigger the Maillard reaction, a complex chemical process that generates ACR as a byproduct. As a result, a wide range of commonly consumed foods can contain ACR residues. ACR is a highly reactive organic chemical that polymerizes and finds use in a variety of industries [17-19]. ACR is frequently utilized in the cosmetics industry and molecular biology research for wastewater treatment [20]. It can be ingested or be exposed to us through our diet or our surroundings, both of which are mentioned further below [21-22].Figure 01 Different sources of exposure to Acrylamide.

# Dietary Exposure: -

The occurrence of ACR in our regular meals or foods is a main source of concern. Diet plays a significant role in ACR exposure for the general population. Consuming foods like French fries, potato chips, bread, cookies, and coffee, all of which can contain notable ACR levels, contributes



to daily ACR intake. Additionally, exposure through dietary supplements and other sources must be considered when assessing ACR toxicity. Overheating, pH, water content, and the reactivity of the different components can all contribute to the formation of ACR. [20] Elevated temperatures increase the formation of ACR along with the time of heating separately or jointly. ACR is formed at temperatures of 190 degrees or higher, as evidenced by the rise in ACR concentrations in French fries after cooking [23]. ACR is found in breast milk, and up to 50% of it is transferred from a pregnant mother to the growing foetus through the placenta [24].

#### **Environmental Exposure**

Environmental ACR primarily originates from industrial processes, including the production of plastics, wastewater treatment, cosmetics and textile industries, cigarette smoke, and other sources. It is used as an intermediate in organic compound manufacturing and as a monomer in the production of polyacrylamide, a polymer widely employed in various industries. ACR can also be found in the waste streams of these processes, potentially contaminating soil and water [25,26] ACR can be ingested, inhaled, or comes into contact with the skin. A single cigarette includes around 1  $\mu$ g of ACR. Snuff, tobacco strips, and other tobacco products contain ACR in the range of 100 to 367 ng/g dry weight [27].

#### **Occupational exposure: -**

Inhalation or direct contact with skin or mucous membranes are the most common methods

of occupational exposure. Furthermore, some personal care products (for example, cosmetics) include free ACR, which can be absorbed through dermal contact; however, estimating dermal exposure is challenging [28].

#### Tobacco exposure: -

In tabaco, more than 8,000 compounds have been discovered, with more than 60 of them being recognized to be carcinogenic [29]. Hazardous substances were tested in both mainstem and side stream smoke. Tobacco pyrolysis also produces ACR, and tobacco use is a major source of ACR exposure [30].

#### Acrylamide in Processed Food:

Acrylamide in food is mostly formed during baking and frying by heat-induced interactions between the amino group of the free amino acid asparagine and the carbonyl group of reducing sugars such as glucose. Plant-based foods high in both precursors include potatoes and cereals (barley, rice, and wheat), but not animal meals like chicken, meat, and fish. High-acrylamide processed foods include French fries, potato chips, tortilla chips, bread crust, crispbread, and many baked products and cereal formulations However, the observed wide variations in acrylamide levels in different food categories as well as different brands of the same food category (e.g., French fries, potato chips) appear to be due to changes in processing circumstances (e.g., temperature, time, nature of frving oil, nature of food matrix). Table 02 shows the Content of Acrylamide in Food Products

THE PRODUCTS TYPE	THE ACRYLAMIDE CONTENT
	(µG/KG)
Bread (rolls, bread, bagels)	70-430
Potato chips	<50-3500
Potato fries	200-2287
Boiled potatoes	48
Cookies, crackers, biscuits	<30-3200
Rusks	80-1200
Cereals	30-1400
Gingerbread cookies	<50-100
Chocolate (powder)	64-457
Nuts, peanut butter	64-457
Meat, poultry	30-64
Baked asparagus	143

 Table 01: - The Content of Acrylamide in Food Products:

 E PRODUCTS TYPE
 THE
 ACRYLAMIDE
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# ACR and oxidative stress

In the pathology of chemical-induced neurodegeneration, oxidative stress is a critical event. When the rate of production of free oxygen radicals (ROS) exceeds the rate of neutralization, oxidative stress occurs. Excessive free radicals can cause biological molecules to oxidize, such as lipid peroxidation, enzyme oxidation, and DNA base oxidation. This results in cell organelle damage, slowed cell metabolism, DNA fragmentation, and cell death [31-33]. According to some experimental evidence, it also shown that ACR-induced oxidative stress is characterized by increased lipid peroxidation (LPO) and protein carbonyl content, as well as a decrease in enzymic and non-enzymic antioxidants[34]. Studies also investigate the negative effects of ACR on PC12 cells. [35].In the brain tissues of rats exposed to 40 mg/kg ACR for four weeks, the level of MDA (an important and biomarker of oxidative stress lipid peroxidation) increased significantly, while the content of GSH (a biologically important intracellular thiol acting as a free radical scavenger) and the activities of SOD and GSH-Px (two important antioxidant enzymes) decreased significantly [36].

# 1.6 ACR and mitochondrial dysfunction

Mitochondria are the organelles that generate 90 percent of cellular ATP. The function of mitochondria is especially important in nerve cells specified for ATP generation [37]. The neurons are specific cell which requires the higher energy to support the cell activities, especially in the synaptic region. Mitochondria is the main site for ROS production, which is involved normally under physiological conditions, that is mainly produced at the site of mitochondrial enzyme complex I and III in the respiratory chain of mitochondria. Some studies reported that the mitochondrial complexes II are also involved in ROS production [38 - 40]. ACR showed a concentration-dependent reduction in cell viability and induced apoptosis. ACR toxicity was also observed due to impairment in the mitochondrial respiratory chain, aerobic glycolysis, and lower expression of the complex I, III, and IV subunits. The activation of mitochondrion-driven apoptotic signalling is triggered by a decrease in mitochondrial membrane potential and the ratio of Bcl-2/Bax. Increased NF-kB expression, as well as downstream inducible nitric oxide synthase (iNOS) and nitric oxide production, suggest that ACR has a pro-inflammatory effect.[41] ACR-induced

mitochondria-dependent apoptosis was mediated by the mitogen-activated protein kinases (MAPK) and Nrf2 signalling pathways [42]. [43-45]. Currently ACR-induced cellular toxicity has been shown to inhibit proliferation and differentiation, activate mitochondrial-mediated apoptosis, and downregulate antioxidant signalling pathways in neurons [46- 48].

# Neurotransmitter and neurotoxicity

ACR causes neurotoxicity by covalently binding to critical pre-synaptic protein thiol groups, resulting in a reduction in neurotransmitter release. Furthermore, ACR has been shown to affect DA receptor density, DA uptake, and dopamine release [49-50], some studies have shown that considerable decrease in the neurotransmitter's dopamine and norepinephrine in the brains of rats and zebrafish. [51-52].Some studies also revealed that the increased AChE activity inhibits cell proliferation and promotes apoptosis. ACR caused a significant increase in neurotransmission markers in the brain, such as AChE activity [53].

# Neuromodulator alteration&Inflammation

Upregulation of glial fibrillary acidic protein (GFAP), an astrocytic marker, has been reported in all types of central nervous system injury. Exposure to be ACR increase the expression of GFAP [54] BDNF is an important neurotrophic brain development factor that promotes neuron growth, synaptic function, and neural plasticity during brain development. The most important functions of BDNF are neuron protection from toxic effects and neuron survival [55]. A current study has also discovered that giving ACR to pregnant rats reduces foetal brain BDNF levels [56-58].

# Role of berberine in various disease: -

Berberine is a yellow alkaloid which occurs in numerous plants [59, 60]. Berberine is the principal component for many popular medicinal plants, such as Coptidis chinensis Franch. (family Ranunculaceae), PhellodendronchinenseSchneid. (family Rutaceae), and Mahonia bealei (Fort.) Carr. (family Berberidaceae) [62].BBR was found to be a small molecule with a molecular weight of only 371.8 Da [63]. BBR has been used clinically to treat bacterial diarrhea, hypercholesterolemia, type 2 diabetes, cardiac disease, cancer, and more [63-68]. Several therapeutic effects of berberine have been identified against cancer, obesity, congestive heart failure,[69] inflammation, atherosclerosis,



neurodegenerative diseases,[70] rheumatoid arthritis, cardiovascular diseases,[71] and metabolic disorders, such as dyslipidemia, impaired fasting glucose, metabolic syndrome, and diabetes [69,72-74].Historically, berberine has also been used as a yellow dye, due to its yellow color.In tradition, the main activity of berberine has been evidenced to possess antimicrobial properties against various bacteria, fungi, protozoans, helminths, chlamydia, and viruses [75-78].Berberine has also been evaluated as a treatment for hypercholesterolemia, via reducing serum triglycerides, cholesterol, and low-density lipoprotein (LDL) cholesterol [79-81] Multiple studies have indicated that berberine has been found to be beneficial to cancer,[82] obesity, atherosclerosis.[83] rheumatoid arthritis. cerebrovascular diseases, fever, headaches, high blood pressure, immune system, irritable bowel syndrome, leukemia, leukopenia, liver disease (alcoholic), osteoporosis and respiratory disorders.

# Therapeutic role of Berberine on neurotoxicity

Berberine has been found to possess multiple neuroprotective effects and improve survival, development, and function of neurons, while protecting these electrically excitable brain cells[84] Furthermore, there has been strong evidence that berberine has a close relationship with neurodegenerative diseases,[85] includingAlzheimer's disease(AD),[86] Parkinson's disease (PD)[87] and Huntington's disease(HD) [88].

# II. CONCLUSION

In conclusion, this review highlights berberine's excellent neuroprotective ability against neurotoxicity generated by acrylamide. The comprehensive analysis of existing literature reveals that berberine exerts its protective effects through multifaceted mechanisms, including the attenuation of oxidative stress, modulation of apoptosis, and suppression of inflammatory responses in neural tissues. The observed neuroprotective properties position berberine as a promising candidate for mitigating the detrimental impact of Acrylamide on the nervous system. These insights not only contribute to the understanding of berberine's pharmacological actions but also highlight its potential therapeutic value in preventing or ameliorating neurotoxicity associated with Acrylamide exposure, warranting further exploration in clinical contexts.

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# Credit author statement

Jitesh Kumar Patwa - wrote the manuscript. Jyoti Yadav, Sanskar Gupta, Ritesh Kumar, Khemendra Chaturvedi – Prepare diagrams and referencing. Ritesh Jain– Design the layout and Kuleshwar Sahu critically revise the manuscript.

# **Conflict of interest**

The author declares that there are no conflicts of interest regarding the publication of this manuscripts.

# Ethical approval

This article does not contain any studies with human participants or animals performed by any of the author.

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# List of abbreviation:

- BBR :- Berberine
- AD :- Alzheimer's disease
- PD :- Parkinson's disease
- ROS :- Reactive oxygen species
- ACR :- Acrylamide
- SOD :- Superoxide dismutase
- LPO: Lipid peroxidation
- WHO: World Health Organization

# **REFERENCES: -**

- [1]. Parng C, Roy NM, Ton C, Lin Y, McGrath P. Neurotoxicity assessment using zebrafish. Journal of pharmacological and toxicological methods. 2007 Jan 1;55(1):103-12.
- [2]. ACR WH. Environmental Health Criteria 49. Geneva: World Health Organization, 1985.
- [3]. Mulloy KB. Two case reports of neurological disease in coal mine preparation plant workers. American journal of industrial medicine. 1996 Jul;30(1):56-61.
- [4]. EFSA Panel on Contaminants in the Food Chain (CONTAM). Scientific opinion on ACR in food. Efsa Journal. 2015 Jun;13(6):4104.



- [5]. JECFA W. Joint FAO/WHO expert committee on food additives. Evaluation of certain food additives and contaminants. Sixty Seventh Report, series. 2006;940:10-5.
- [6]. Friedman M. Chemistry, biochemistry, and safety of ACR. A review. Journal of agricultural and food chemistry. 2003 Jul 30;51(16):4504-26.
- [7]. Smith EA, Oehme FW. ACR and polyACR: a review of production, use, environmental fate and neurotoxicity. Reviews on environmental health. 1991 Oct 1;9(4):215-28.
- [8]. Tilson HA. The neurotoxicity of ACR: an overview. Neurobehavioral toxicology and teratology. 1981 Jan 1;3(4):445-61.
- [9]. Biedermann M, Biedermann-Brem S, Noti A, GROB K. Methods for determining the potential of ACR formation and its elimination in raw materials for food preparation, such as potatoes. MitteilungenausLebensmitteluntersuchung und Hygiene. 2002;93(6):653-67.
- [10]. Tareke E, Rydberg P, Karlsson P, Eriksson S, Törnqvist M. Analysis of ACR, a carcinogen formed in heated foodstuffs. Journal of agricultural and food chemistry. 2002 Aug 14;50(17):4998-5006.
- [11]. Dybing E, Farmer PB, Andersen M, Fennell TR, Lalljie SP, Müller DJ, Olin S, Petersen BJ, Schlatter J, Scholz G, Scimeca JA. Human exposure and internal dose assessments of ACR in food. Food and Chemical Toxicology. 2005 Mar 1;43(3):365-410.
- [12]. FAO., World Health Organization. Food World Safety Programme, Health Organization, World Health Organisation Staff, Joint FAO/WHO Consultation on Health Implications of ACR in Food, Food Programme, WHO. Safety Health Implications of ACR in Food: Report of a Joint FAO/WHO Consultation, WHO Headquarters, Geneva, Switzerland, 25-27 June 2002. World Health Organization; 2002 Dec 28.
- [13]. Moldoveanu SC, Gerardi AR. ACR analysis in tobacco, alternative tobacco products, and cigarette smoke. Journal of chromatographic science. 2011 Mar 1;49(3):234-42.

- [14]. Erkekoglu P, Baydar T. ACR neurotoxicity. Nutritional neuroscience. 2014 Feb 1;17(2):49-57.
- [15]. Johnson KA, Gorzinski SJ, Bodner KM, Campbell RA, Wolf CH, Friedman MA, Mast RW. Chronic toxicity and oncogenicity study on ACR incorporated in the drinking water of Fischer 344 rats. Toxicology and applied pharmacology. 1986 Sep 15;85(2):154-68.
- [16]. Exon JH. A review of the toxicology of ACR. Journal of Toxicology and Environmental Health, Part B. 2006 Dec 1;9(5):397-412.
- [17]. Mottram DS, Wedzicha BL, Dodson AT. Acrylamide is formed in the Maillard reaction. Nature. 2002 Oct 3;419(6906):448-9.
- [18]. Raffan S, Halford NG. ACR in food: Progress in and prospects for genetic and agronomic solutions. Annals of Applied Biology. 2019 Nov;175(3):259-81.
- [19]. Koszucka A, Nowak A, Nowak I, Motyl I. ACR in human diet, its metabolism, toxicity, inactivation and the associated European Union legal regulations in food industry. Critical reviews in food science and nutrition. 2020 May 30;60(10):1677-92.
- [20]. Pedreschi F, Mariotti MS, Granby K. Current issues in dietary ACR: formation, mitigation and risk assessment. Journal of the Science of Food and Agriculture. 2014 Jan 15;94(1):9-20.
- [21]. Kumar J, Das S, Teoh SL. Dietary ACR and the risks of developing cancer: Facts to ponder. Frontiers in nutrition. 2018 Feb 28;5:14.
- [22]. Pennisi M, Malaguarnera G, Puglisi V, Vinciguerra L, Vacante M, Malaguarnera M. Neurotoxicity of ACR in exposed workers. International journal of environmental research and public health. 2013 Sep;10(9):3843-54.
- [23]. Chen MJ, Hsu HT, Lin CL, Ju WY. A statistical regression model for the estimation of ACR concentrations in French fries for excess lifetime cancer risk assessment. Food and Chemical Toxicology. 2012 Oct 1;50(10):3867-76
- [24]. Annola K, Karttunen V, Keski-Rahkonen P, Myllynen P, Segerbäck D, Heinonen S, Vähäkangas K. Transplacental transfer of ACR and glycidamide are comparable to



that of antipyrine in perfused human placenta. Toxicology letters. 2008 Nov 10;182(1-3):50-6.

- [25]. Smith EA, Oehme FW. Acrylamide and polyacrylamide: a review of production, use, environmental fate and neurotoxicity. Reviews on environmental health. 1991 Oct;9(4):215
- [26]. Hang B, Wang P, Zhao Y, Chang H, Mao JH, Snijders AM. Thirdhand smoke: Genotoxicity and carcinogenic potential. Chronic diseases and translational medicine. 2020 Mar 1;6(1):27-34.
- [27]. Semla M, Goc Z, Martiniaková M, Omelka R, Formicki G. ACR: a common food toxin related to physiological functions and health. Physiological research. 2017 Apr 1;66(2).
- [28]. ObónSantacana M. Dietary intake and biomarkers of ACR exposure and risk of endometrial and ovarian cancer: A molecular epidemiologic study in the European Prospective Investigation into Cancer and Nutrition
- [29]. Hecht SS, Szabo E. Fifty years of tobacco carcinogenesis research: from mechanisms to early detection and prevention of lung cancer. Cancer prevention research. 2014 Jan 1;7(1):1-8.
- [30]. Hirvonen T, Kontto J, Jestoi M, Valsta L, Peltonen K, Pietinen P, Virtanen SM, Sinkko H, Kronberg-Kippilä C, Albanes D, Virtamo J. Dietary ACR intake and the risk of cancer among Finnish male smokers. Cancer Causes & Control. 2010 Dec;21(12):2223-9.
- [31]. ObónSantacana M. Dietary intake and biomarkers of ACR exposure and risk of endometrial and ovarian cancer: A molecular epidemiologic study in the European Prospective Investigation into Cancer and Nutrition.
- [32]. Rahman T, Hosen I, Islam MT, Shekhar HU. Oxidative stress and human health.
- [33]. Sahu K, Langeh U, Singh C, Singh A. Crosstalk between anticancer drugs and mitochondrial functions. Current Research in Pharmacology and Drug Discovery. 2021 Jan 1;2:100047.
- [34]. Selvakumar K, Bavithra S, Ganesh L, Krishnamoorthy G, Venkataraman P, Arunakaran J. Polychlorinated biphenyls induced oxidative stress mediated neurodegeneration in hippocampus and

behavioral changes of adult rats: Anxiolytic-like effects of quercetin. Toxicology letters. 2013 Sep 12;222(1):45-54.

- [35]. Ren X, Zou L, Zhang X, Branco V, Wang J, Carvalho C, Holmgren A, Lu J. Redox signaling mediated by thioredoxin and glutathione systems in the central nervous system. Antioxidants & redox signaling. 2017 Nov 1;27(13):989-1010.
- [36]. ZHAO, Mengyao, et al. Acrylamideinduced neurotoxicity in primary astrocytes and microglia: roles of the Nrf2-ARE and NF-κB pathways. Food and chemical toxicology, 2017, 106: 25-35.
- [37]. GUO, Jie, et al. The anti-apoptotic, antioxidant and anti-inflammatory effects of curcumin on acrylamide-induced neurotoxicity in rats. BMC Pharmacology and Toxicology, 2020, 21.1: 1-10.
- [38]. Ren X, Zou L, Zhang X, Branco V, Wang J, Carvalho C, Holmgren A, Lu J. Redox signaling mediated by thioredoxin and glutathione systems in the central nervous system. Antioxidants & redox signaling. 2017 Nov 1;27(13):989-1010.
- [39]. Lobo V, Patil A, Phatak A, Chandra N. Free radicals, antioxidants and functional foods: Impact on human health. Pharmacognosy reviews. 2010 Jul;4(8):118
- [40]. Nissanka N, Moraes CT. Mitochondrial DNA damage and reactive oxygen species in neurodegenerative disease. FEBS letters. 2018 Mar;592(5):728-42.
- [41]. Brand MD, Nicholls DG. Assessing mitochondrial dysfunction in cells. Biochemical Journal. 2011;435(2):297-312.
- [42]. Chiswick BR, Miller PW. International migration and the economics of language. Handbook of the economics of international migration. 1: Elsevier; 2015. p. 211-69.
- [43]. Quinlan CL, Orr AL, Perevoshchikova IV, Treberg JR, Ackrell BA, Brand MD. Mitochondrial complex II can generate reactive oxygen species at high rates in both the forward and reverse reactions. Journal of Biological Chemistry. 2012;287(32):27255-6
- [44]. Olianas MC, Dedoni S, Onali P. Involvement of store-operated Ca2+ entry



in activation of AMP-activated protein kinase and stimulation of glucose uptake by M3 muscarinic Ach receptors in human neuroblastoma cells. Biochimica et Biophysica Acta (BBA)-Molecular Cell Research. 2014 Dec 1;1843(12):3004-17.

- [45]. Zhao M, Wang FS, Hu X, Chen F, Chan HM. ACR-induced neurotoxicity in primary astrocytes and microglia: roles of the Nrf2-ARE and NF-κB pathways. Food and chemical toxicology. 2017 Aug 1;106:25-35.
- [46]. Teismann P. Schulz JB. Cellular pathology of Parkinson's disease: astrocytes, microglia and inflammation. and tissue research. 2004 Cell Oct;318(1):149-61
- [47]. Santhanasabapathy R, Vasudevan S, Anupriya K, Pabitha R, Sudhandiran G. Farnesol quells oxidative stress, reactive gliosis and inflammation during ACRinduced neurotoxicity: Behavioral and biochemical evidence. Neuroscience. 2015 Nov 12;308:212-
- [48]. MONEIM, Ahmed E. Abdel. The neuroprotective effect of berberine in mercury-induced neurotoxicity in rats. Metabolic brain disease, 2015, 30.4: 935-942.
- [49]. Hagemann TL, Paylor R, Messing A. Deficits in adult neurogenesis, contextual fear conditioning, and spatial learning in a Gfap mutant mouse model of Alexander disease. Journal of Neuroscience. 2013 Nov 20;33(47):18698-706.
- [50]. Kaneko R, Hagiwara N, Leader K, Sueoka N. Glial-specific cAMP response of the glial fibrillary acidic protein gene cell lines. Proceedings of the National Academy of Sciences. 1994 May 10;91(10):4529-33
- [51]. DIXIT, Rakesh, et al. Effect of acrylamide on biogenic amine levels, monoamine oxidase, and cathepsin D activity of rat brain. Environmental research, 1981, 26.1: 168-173.
- [52]. FARIA, Melissa, et al. Acrylamide acute neurotoxicity in adult zebrafish. Scientific reports, 2018, 8.1: 1-14.
- [53]. Owens DF, Kriegstein AR. Is there more to GABA than synaptic inhibition?. Nature Reviews Neuroscience. 2002 Sep;3(9):715-27.

- [54]. Mattson MP, Maudsley S, Martin B. BDNF and 5-HT: a dynamic duo in agerelated neuronal plasticity and neurodegenerative disorders. Trends in neurosciences. 2004 Oct 1;27(10):589-94.
- [55]. Babin PJ, Goizet C, Raldúa D. Zebrafish models of human motor neuron diseases: advantages and limitations. Progress in neurobiology. 2014 Jul 1;118:36-58.
- [56]. Gómez-Canela C, Prats E, Piña B, Tauler R. Assessment of chlorpyrifos toxic effects in zebrafish (Danio rerio) metabolism. Environmental Pollution. 2017 Jan 1;220:1231-43.
- [57]. MONEIM, Ahmed E. Abdel. The neuroprotective effect of berberine in mercury-induced neurotoxicity in rats. Metabolic brain disease, 2015, 30.4: 935-942.
- [58]. Mueller NP, Carloni P, Alfonso-Prieto M. Molecular determinants of acrylamide neurotoxicity through covalent docking. Frontiers in Pharmacology. 2023 Mar 2;14:1125871.
- [59]. Hahn FE, Ciak J. Berberine. InMechanism of action of antimicrobial and antitumor agents 1975 (pp. 577-584). Berlin, Heidelberg: Springer Berlin Heidelberg.
- [60]. Cai Z, Wang C, Yang W. Role of berberine in Alzheimer's disease. Neuropsychiatric disease and treatment. 2016 Oct 3:2509-20.
- [61]. Imanshahidi M, Hosseinzadeh H. Pharmacological and therapeutic effects of Berberis vulgaris and its active constituent, berberine. Phytotherapy research. 2008 Aug;22(8):999-1012.
- [62]. Wang K, Feng X, Chai L, Cao S, Qiu F. The metabolism of berberine and its contribution to the pharmacological effects. Drug metabolism reviews. 2017 Apr 3;49(2):139-57.
- [63]. Kong W, Wei J, Abidi P, Lin M, Inaba S, Li C, Wang Y, Wang Z, Si S, Pan H, Wang S. Berberine is a novel cholesterollowering drug working through a unique mechanism distinct from statins. Nature medicine. 2004 Dec 1;10(12):1344-51.
- [64]. Zhang H, Wei J, Xue R, Wu JD, Zhao W, Wang ZZ, Wang SK, Zhou ZX, Song DQ, Wang YM, Pan HN. Berberine lowers blood glucose in type 2 diabetes mellitus patients through increasing insulin



receptor expression. Metabolism. 2010 Feb 1;59(2):285-92.

- [65]. Li BX, Yang BF, Zhou J, Xu CQ, Li YR. Inhibitory effects of berberine on IK1, IK, and HERG channels of cardiac myocytes. Acta PharmacologicaSinica. 2001 Feb 1;22(2):125-31.
- [66]. Jiang Q, Liu P, Wu X, Liu W, Shen X, Lan T, Xu S, Peng J, Xie X, Huang H. Berberine attenuates lipopolysaccharideinduced extracelluar matrix accumulation and inflammation in rat mesangial cells: involvement of NF-κBsignaling pathway. Molecular and cellular endocrinology. 2011 Jan 1;331(1):34-40.
- [67]. Hayashi K, Minoda K, Nagaoka Y, Hayashi T, Uesato S. Antiviral activity of berberine and related compounds against human cytomegalovirus. Bioorganic & medicinal chemistry letters. 2007 Mar 15;17(6):1562-4.
- [68]. Iizuka N, Miyamoto K, Okita K, Tangoku A, Hayashi H, Yosino S, Abe T, Morioka T, Hazama S, Oka M. Inhibitory effect of CoptidisRhizoma and berberine on the proliferation of human esophageal cancer cell lines. Cancer letters. 2000 Jan 1;148(1):19-25.
- [69]. Chang W, Li K, Guan F, Yao F, Yu Y, Zhang M, Hatch GM, Chen L. Berberine pretreatment confers cardioprotection against ischemia–reperfusion injury in a rat model of type 2 diabetes. Journal of cardiovascular pharmacology and therapeutics. 2016 Sep;21(5):486-94.
- [70]. Jiang W, Li S, Li X. Therapeutic potential of berberine against neurodegenerative diseases. Science China Life Sciences. 2015 Jun;58:564-9.
- [71]. Jin Y, Khadka DB, Cho WJ. Pharmacological effects of berberine and its derivatives: a patent update. Expert Opinion on Therapeutic Patents. 2016 Feb 1;26(2):229-43.
- [72]. Caliceti C, Franco P, Spinozzi S, Roda A, FG Cicero A. Berberine: new insights from pharmacological aspects to clinical evidences in the management of metabolic disorders. Current Medicinal Chemistry. 2016 Apr 1;23(14):1460-76.
- [73]. Pirillo A, Catapano AL. Berberine, a plant alkaloid with lipid-and glucose-lowering properties: from in vitro evidence to

clinical studies. Atherosclerosis. 2015 Dec 1;243(2):449-61.

- [74]. Chang W, Zhang M, Meng Z, Yu Y, Yao F, Hatch GM, Chen L. Berberine treatment prevents cardiac dysfunction and remodeling through activation of 5'-adenosine monophosphate-activated protein kinase in type 2 diabetic rats and in palmitate-induced hypertrophic H9c2 cells. European journal of pharmacology. 2015 Dec 15;769:55-63.
- [75]. Dziedzic A, Wojtyczka RD, Kubina R. Inhibition of oral streptococci growth induced by the complementary action of berberine chloride and antibacterial compounds. Molecules. 2015 Jul 28;20(8):13705-24.
- Wojtyczka RD, Dziedzic A, Kępa M, [76]. Kubina R, Kabała-Dzik A, Mularz T, Berberine Idzik D. enhances the antibacterial activity of selected antibiotics against coagulase-negative Staphylococcus strains in vitro. Molecules. 2014 May 22;19(5):6583-96.
- [77]. Harikumar KB, Kuttan G, Kuttan R. Inhibition of progression of erythroleukemia induced by Friend virus in BALB/c mice by natural products-Berberine, Curcumin and Picroliv. Journal of Experimental Therapeutics & Oncology. 2008 Dec 1;7(4).
- [78]. Schmeller T, Latz-Brüning B, Wink M. Biochemical activities of berberine, palmatine and sanguinarine mediating chemical defence against microorganisms and herbivores. Phytochemistry. 1997 Jan 1;44(2):257-66.
- [79]. Ghareeb DA, Khalil S, Hafez HS, Bajorath J, Ahmed HE, Sarhan E, Elwakeel E, El-Demellawy MA. Berberine reduces neurotoxicity related to nonalcoholic steatohepatitis in rats. Evidence-Based Complementary and Alternative Medicine. 2015 Jan 1;2015.
- [80]. Dong H, Zhao Y, Zhao L, Lu F. The effects of berberine on blood lipids: a systemic review and meta-analysis of randomized controlled trials. Planta medica. 2013 Apr;79(06):437-46.
- [81]. Meng S, Wang LS, Huang ZQ, Zhou Q, Sun YG, Cao JT, Li YG, Wang CQ. Berberine ameliorates inflammation in patients with acute coronary syndrome following percutaneous coronary



intervention. Clinical and Experimental Pharmacology and Physiology. 2012 May;39(5):406-11.

- [82]. Naveen CR, Gaikwad S, Agrawal-Rajput R. Berberine induces neuronal differentiation through inhibition of cancer stemness and epithelial-mesenchymal transition in neuroblastoma cells. Phytomedicine. 2016 Jun 15;23(7):736-44.
- [83]. Li H, He C, Wang J, Li X, Yang Z, Sun X, Fang L, Liu N. Berberine activates peroxisome proliferator-activated receptor gamma to increase atherosclerotic plaque stability in Apoe-/mice with hyperhomocysteinemia. Journal of Diabetes Investigation. 2016 Nov;7(6):824-32.
- [84]. Kysenius K, Huttunen HJ. Stress-induced upregulation of VLDL receptor alters Wnt-signaling in neurons. Experimental cell research. 2016 Jan 15;340(2):238-47.
- [85]. Zhang J, Yang JQ, He BC, Zhou QX, Yu HR, Tang Y, Liu BZ. Berberine and total base from rhizomacoptis chinensis attenuate brain injury in an aluminuminduced rat model of neurodegenerative

disease. Saudi medical journal. 2009 Jun 1;30(6):760-6.

- [86]. Hussien HM, Abd-Elmegied A, Ghareeb DA, Hafez HS, Ahmed HE, Abd Elmoneam N. Neuroprotective effect of berberine against environmental heavy metals-induced neurotoxicity and Alzheimer's-like disease in rats. Food and chemical toxicology. 2018 Jan 1;111:432-44.
- [87]. Kim M, Cho KH, Shin MS, Lee JM, Cho HS, Kim CJ, Shin DH, Yang HJ. Berberine prevents nigrostriatal dopaminergic neuronal loss and suppresses hippocampal apoptosis in mice with Parkinson's disease. International Journal of Molecular Medicine. 2014 Apr 1;33(4):870-8.
- [88]. Jiang W, Wei W, Gaertig MA, Li S, Li XJ. Therapeutic effect of berberine on Huntington's disease transgenic mouse model. PloS one. 2015 Jul 30;10(7):e0134142